

# Approach to Neurotuberculosis the Great Chameleon

Dr Mona Thakre  
Consultant Neurologist  
MD, DM, DNB, SCE(UK)Neurology,FAAN  
AL Zahra Hospital ,Dubai, UAE  
E mail- [monathakre@yahoo.com](mailto:monathakre@yahoo.com)

# Disclosures

None

# Learning Objectives

- Understanding global TB situation with special emphasis on MDR/XDR TB
- Understanding various CNS presentations of TB
- Learning the latest and recommended investigations
- Understanding the treatment guidelines for CNS TB including MDR/XDR TB

# Case history

BJ 25yrs, Female

fever, cough —diagnosed right Pleural effusion secondary to TB started on AKT4 - 1 month

Investigation done there Pleural fluid —No org

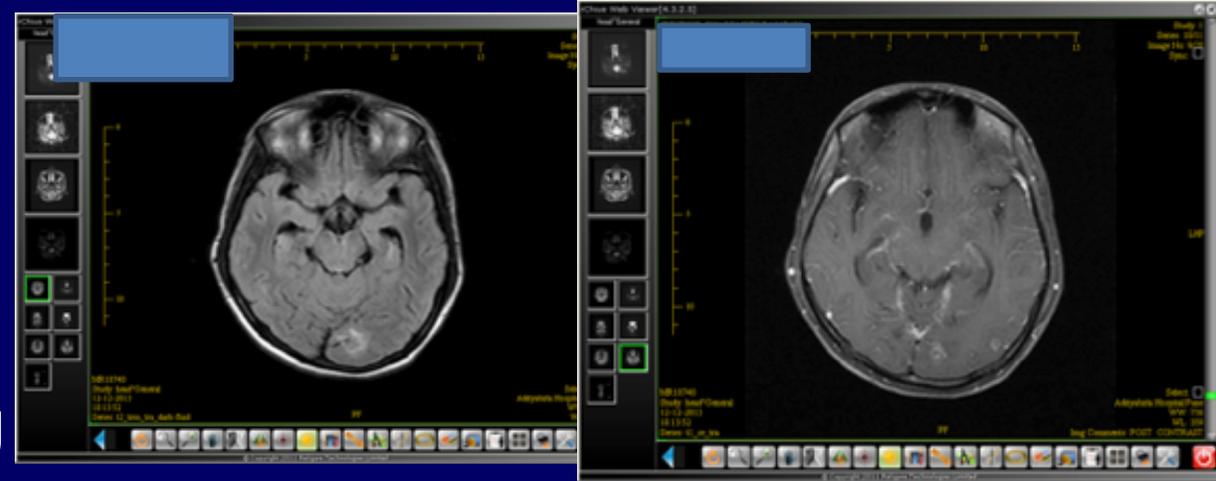
Sputum 3 samples – AFB negative

Developed Altered Sensorium ,CT brain – N

Shifted to our hospital ,Conscious , febrile.

Confused, Neck stiffness

Chest Xray military mottling, MRI brain – Left Occipital ring enhancing lesion with meningeal enhancement



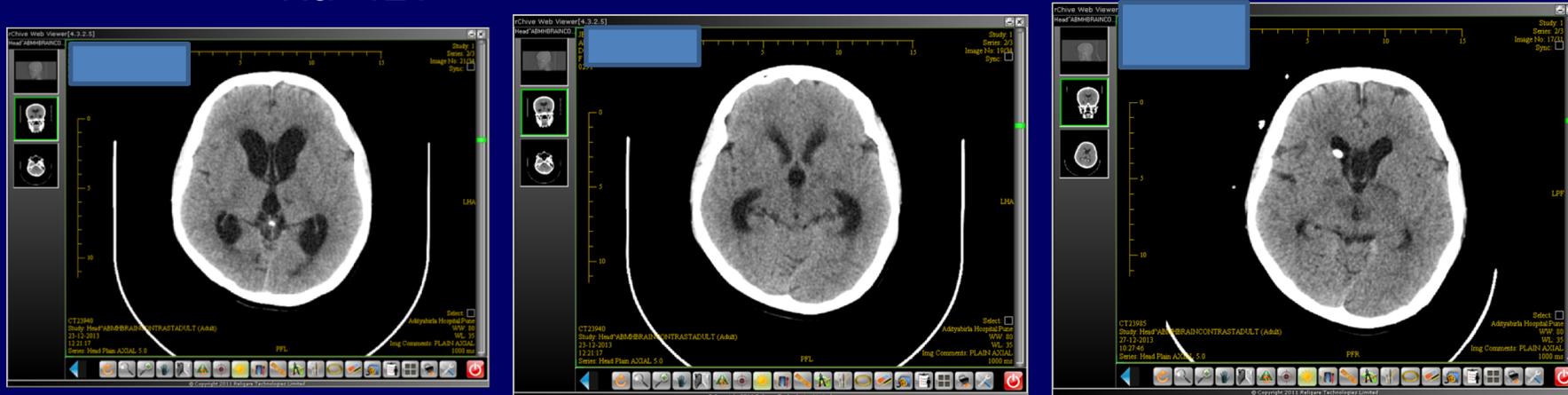
CSF- Protein- 159

- Sugar - <20 ( parallel blood sugar  
90mg/dl)

- TLC – 190 90% lymphocytes

- ADA- 82, CSF- NO AFB, India Ink- neg, sent for C&S

- Na -121



6 week later CSF culture : Mycobacterium Tuberculosis Complex  
Sensitivity-

Ethambutol- Resistant

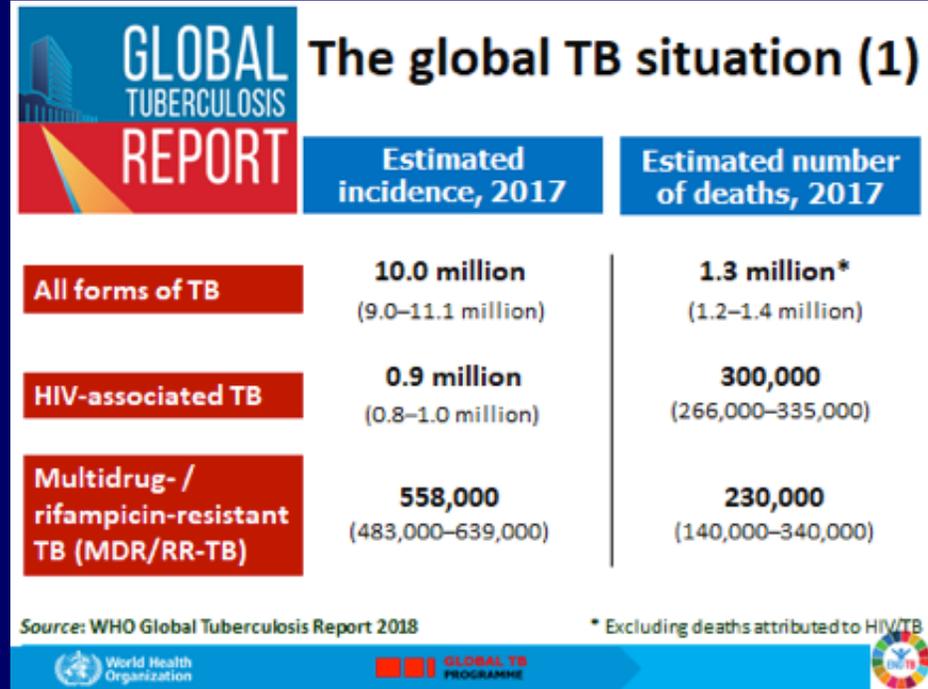
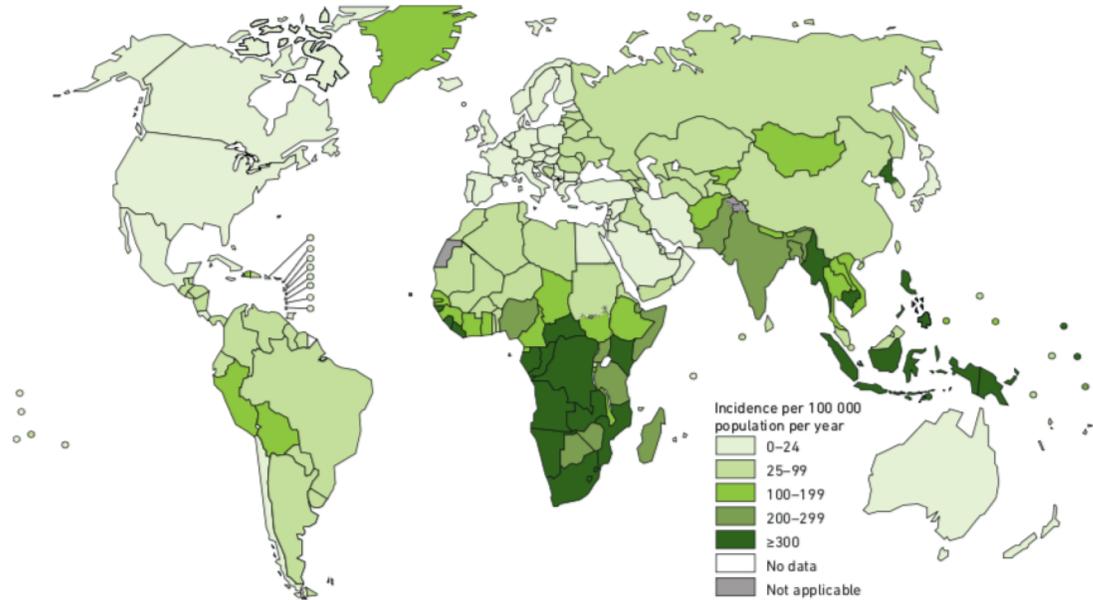
Pyrazinamide- Resistant

Rifampicin - Resistant

Isoniazid --- Resistant



Estimated TB incidence rates, 2017



Incidence of TB in the UAE is considered low at 0.79 per 100,000 inhabitants compared to other countries, down from 4.2 per 100,000 in 2000, according to the Global Tuberculosis Report.

- CNS tuberculosis occurs in up to 10% and has protean clinical manifestations. The burden is directly proportional to the prevalence of TB

Tuberculous meningitis is the most devastating form of extra-pulmonary tuberculosis with 30% mortality and disabling neurological sequelae in > 25% survivors.

# Causative organism

- CNS tuberculosis is caused by the human strain of Mycobacterium tuberculosis.
- However in immunocompromised patients, atypical mycobacteria are an important cause of infection.
- They are now called non-tuberculous mycobacteria which include:
  - Mycobacterium avium
  - Mycobacterium intracellulare



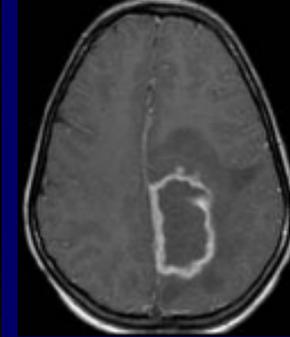
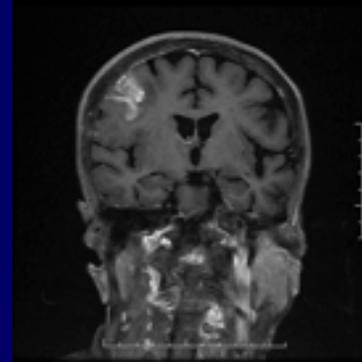
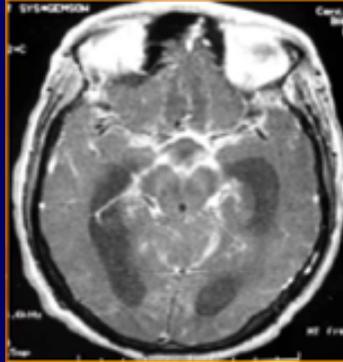
# Pathophysiology

- CNS tuberculosis is secondary to disease elsewhere in the body and reach the brain by hematogenous route.
- Initial small tuberculous lesions (Rich foci) develop in meninges, subpial or subependymal surface of the brain or the spinal cord, and may remain dormant for years.
- Reactivation may be due to endogenous factors: Innate immunological and non immunological defenses ,level of function of cell mediated immunity.
- Release of M tuberculosis results in a T lymphocyte dependent necrotising granulomatous inflammatory response.
- Thick gelatinous exudate around the sylvian fissures, basal cisterns, brainstem and cerebellum causing
  - Hydrocephalous
  - Adhesive arachnoiditis
  - Obliterative vasculitis

# Classification and presentation

- Intracranial

- Tuberculous meningitis
- Tuberculoma
- Tuberculous abscess
- Tuberculous encephalopathy
- Tuberculous vasculopathy



- Spinal

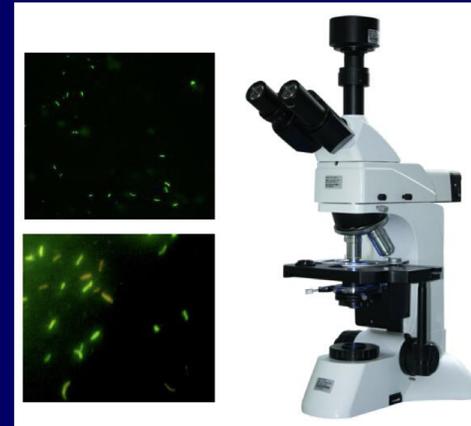
- Pott's spine and Pott's paraplegia
- Tuberculous arachnoiditis
- Spinal tuberculoma
- Spinal meningitis



# Cerebrospinal fluid examination

- Predominantly lymphocytic pleocytosis, increased proteins, low CSF/ blood glucose ratio.
- WBC count can be normal in presence of depressed CMI (elderly and HIV positive individuals).
- Smear is +ve in 10%, can be increased by examining large volume of CSF.
- Culture is +ve in 25-70%. Repeat CSF frequently shows a falling glucose level, a rising protein concentration and a shift to mononuclear predominance. CSF cell counts decrease by 50% during the first month but may not become normal for a year.
- CSF glucose becomes normal in 1 to 2 months and protein becomes normal by 12 months or longer.
- CSF cultures should be sterile by the first month, but PCR results may remain positive for a month
- Gene expert -centrifugation of larger CSF volumes (median: 6 mL, interquartile range: 4–10 mL) resulted in 72% (13/18) sensitivity, which was equivalent to culture (71%, 12/17).

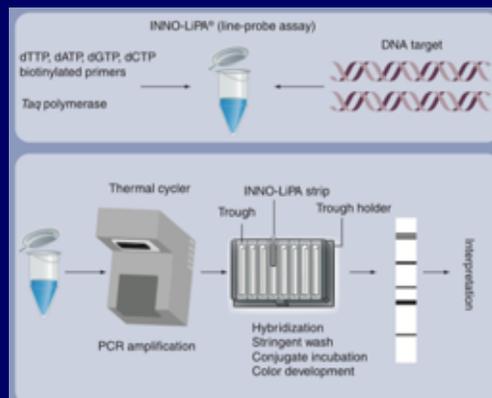
| DIAGNOSTIC PLATFORM      | TEST NAME  | TURNAROUND TIME                                   | DESCRIPTION AND COMMENTS  |
|--------------------------|--|---|---|
| Smear microscopy         | Conventional light microscopy – Ziehl-Neelsen      | 2 hours   | Less sensitive than fluorescent/LED microscopy.   |
|                          | Conventional fluorescence microscopy               |   | Requires a quartz-halogen or high-pressure mercury vapour lamp. Sensitivity is improved over light microscopy, observation time is reduced. Expensive.  |
|                          | Light emitting diode (LED) fluorescence microscopy |   | LED microscopes improve sensitivity by 10% over conventional light microscopy. Observation time is similar to conventional fluorescence microscopy.<br>LED conversion kits for light microscopes are available. |
| Solid culture            | Lowenstein–Jensen                                  | 3 weeks smear positive                            | Egg-based medium, inexpensive.  |
|                          | Middlebrook and Cohn 7H10                          | 4–8 weeks smear negative                          | Agar based medium. Less prone to contamination than Lowenstein–Jensen but more expensive.   |
| Automated liquid culture |  | 8 days smear positive<br>2–6 weeks smear negative | Liquid culture systems. Fully automated systems that use either fluorimetric or colourimetric detection.  |



| DIAGNOSTIC PLATFORM                                    | TEST NAME  | TURNAROUND TIME                                  | DESCRIPTION AND COMMENTS   |
|--|--|--|--|
| Non-commercial WHO endorsed culture and DST techniques | Media-based microscopic observation drug susceptibility (MODS) | 2–21 days direct<br>3–4 weeks indirect           | MODS is a manual liquid technique that uses basic laboratory equipment (including an inverted microscope). Colonies are observed through the bottom of a sealed plastic container. Allows for H and R DST. MODS requires additional staff skills and a containment laboratory.   |
|  | Nitrate reductase assay (NRA)                                  | 6–9 days direct<br>7–11 weeks indirect           | NRA is a colourimetric test using solid media. Allows for H and R DST. TB cells are cultured for 10 days and Greiss reagent is added, which indicates the presence of growing cells.   |
|  | Colourimetric redox indicator (CRI)                            | 3–5 weeks  | CRI is an indirect colourimetric test using liquid media. TB cell are cultured in the presence of a dye. Allows for H and R DST.   |
| Molecular testing                                      | Line probe assay (LPA)   | 1–2 day (direct on smear positive specimen only) | Two LPA have been developed to detect <i>M. tuberculosis</i> resistant to R and H either directly or indirectly. DNA targets are amplified by PCR and hybridized to immobilized oligonucleotide targets. Results are visualized colourimetrically.<br><br>If it is a smear negative specimen, culture must be grown first. |
|  | Xpert MTB/RIF  | 2 hours  | A fully automated test working in a dedicated platform performing detection of MTB and R resistance, using real-time PCR. Results are available in less than two hours.  |

# Drug susceptibility testing

- Phenotypic DST (conventional DST)
- Genotypic DST



## Summary table of accuracy

| Drug | Genes                                 | Accuracy           | Comment   |
|------|---------------------------------------|--------------------|---|
| INH  | katG, inhA, mabA (fabG1)<br>ahpC-oxyR | Se: 84% / Sp: 98%  | All mutations (including not confidence-graded) |
| RIF  | rpoB                                  | Se: 96% / Sp: 99%  |   |
| FQ   | gyrA, gyrB                            | Se: 89% / Sp: 100% |   |
| PZA  | pncA                                  | Se: 83% / Sp: 98%  |   |
| AMK  | eis, rrs                              | Se: 79% / Sp: 100% |   |



# Patterns of drug resistance

- Mono-resistance: resistance to one first-line anti-TB drug only.
- Poly-resistance: resistance to more than one first-line anti-TB drug, other than both isoniazid and rifampicin.
- Multidrug resistance (MDR): resistance to at least both isoniazid and rifampicin.
- Extensive drug resistance (XDR): resistance to any fluoroquinolone, and at least one of three second-line injectable drugs (capreomycin, kanamycin and amikacin), in addition to multidrug resistance.
- Rifampicin resistance (RR): It includes any resistance to rifampicin, in the form of mono-resistance, poly-resistance, MDR or XDR

# WHO recommended drug groups

| GROUP NAME  | ANTI-TB AGENT   |
|---|---|
| <b>Group 1.</b> First-line oral agents  | Isoniazid<br>Rifampicin<br>Ethambutol<br>Pyrazinamide<br>Rifabutin <sup>a</sup><br>Rifapentine <sup>a</sup>                       |
| <b>Group 2.</b> Injectable anti-TB drugs (injectable agents or parental agents) | Streptomycin <sup>b</sup><br>Kanamycin<br>Amikacin<br>Capreomycin   |
| <b>Group 3.</b> Fluoroquinolones (FQs) <sup>d</sup>                             | Levofloxacin<br>Moxifloxacin<br>Gatifloxacin <sup>c</sup>   |
| <b>Group 4.</b> Oral bacteriostatic second-line anti-TB drugs                   | Ethionamide<br>Prothionamide<br>Cycloserine<br>Terizidone <sup>e</sup><br>Para-aminosalicylic acid<br>Para-aminosalicylate sodium |

| GROUP NAME   | ANTI-TB AGENT  |
|--|--|
| <b>Group 5.</b> Anti-TB drugs with limited data on efficacy and/or long term safety in the treatment of drug-resistant TB (This group includes new anti-TB agents) | Bedaquiline<br>Delamanid<br>Linezolid<br>Clofazimine<br>Amoxicillin/ clavulanate<br>Imipenem/cilastatin <sup>f</sup><br>Meropenem <sup>f</sup><br>High-dose isoniazid<br>Thioacetazone <sup>g</sup><br>Clarithromycin <sup>g</sup> |

# Treatment of drug sensitive CNS TB

- CNS tuberculosis is categorized under TB treatment category I by WHO.
- Initial phase therapy ( 2 mths) with isoniazid, rifampicin, pyrazinamide and streptomycin or ethambutol followed by continuation phase (7 mths) with isoniazid and rifampicin.
- The BTS and IDSA/ATS recommend 9-12 months of ATT. Therapy should be extended to 18 months in patients who do not tolerate pyrazinamide.

# General principles of treatment of MDR TB

- The **intensive phase** (i.e. the initial part of treatment during which a Group 2 injectable agent is used) lasts at least **eight months**
- MDR regimens should include at least pyrazinamide, a fluoroquinolone, an injectable anti-TB drug, ethionamide (or prothionamide) and either cycloserine or PAS (para- aminosalicylic acid) if cycloserine cannot be use
- The total length of treatment is expected to be at least **20 months** in most patients not previously treated for MDR-TB .
- **Antiretroviral therapy (ART)** is recommended for all patients with HIV and drug-resistant TB, irrespective of CD4 cell-count, as early as possible (within the first eight weeks) following initiation of the anti-TB treatment
- Adjuvant use of **corticosteroids** has been shown not to increase mortality when the patient is on an effective regimen.& can be beneficial in conditions like severe central nervous system or pericardial involvement

# Key take home messages

- Early diagnosis requires a high index of suspicion.
- Knowledge of newer diagnostics and awareness of MDR /XDR TB  
Helps in better management of the case
- CT or MRI showing basal meningeal enhancement with any degree of hydrocephalus is strongly suggestive of TBM.
- Clinical outcome depends greatly on the stage of disease and correct treatment initiated.

# References

- Guidelines for the programmatic management of drug-resistant tuberculosis. Geneva: World Health Organization; 2011;1–44. ([http://www.who.int/tb/challenges/mdr/programmatic\\_guidelines\\_for\\_mdrtb/en/index.html](http://www.who.int/tb/challenges/mdr/programmatic_guidelines_for_mdrtb/en/index.html), accessed 15 March 2014).
- <https://www.who.int/tb>
- Zuger A. Tuberculosis. In: Scheld WN, Whitley RJ, Marra CM, editors. Infections of Central Nervous System. Philadelphia: Lippincott, 2004. pp. 441-9.
- Kalita J, Misra UK. Tuberculosis Meningitis. In Misra UK, Kalita J (Eds) Diagnosis and Management of Neurological Disorders. Wolter Kluwers Health New Delhi 2011; pp. 145-66
- Thwaites G, et al. J Neurol Neurosurg Psychiatry 2000;68:289-99
- Prasad K, Singh MB. Corticosteroids for managing tuberculous meningitis. Cochrane Database Syst Rev 2008;(1):CD002244
- Thwaites GE et al. J Neurol Neurosurg Psychiatry 2000; 68: 289-99; Lancet Neurol 2005; 4: 160-70
- <https://radiopaedia.org/articles/tuberculosis-intracranial-manifestations>
- [Companion handbook to the WHO guidelines for the programmatic management of drug-resistant tuberculosis](#)